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Role of MHC Class II Genes in the pathogenesis of pemphigoid $\stackrel{\leftrightarrow}{}, \stackrel{\leftrightarrow}{}, \stackrel{\star}{}$

L.R. Zakka ^a, P. Reche ^b, A.R. Ahmed ^{a,*}

^a Center for Blistering Diseases, 70 Parker Hill Avenue, Boston, MA 02120, United States

^b Immunomedicine Group, Department of Immunology, Facultad de Medicina, Universidad Complutense de Madrid, Ave. Complutense S/N, Pabellon 5, Planta IV, Madrid 28040, Spain

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ABSTRACT

Pemphigoid (Pg) is an autoimmune subepidermal blistering disease that affects the elderly population. The phenotype can be Bullous Pemphigoid (BP), which primarily involves the skin, or Mucous Membrane Pemphigoid (MMP), which primarily involves mucus membranes. Ocular Cicatricial Pemphigoid (OCP) and Oral Pemphigoid (OP) are subsets of MMP. The known antigens in BP are Bullous Pemphigoid Antigen 1 (BPAG1, also known as BP230), Bullous Pemphigoid Antigen 2 (BPAG2, also known as BP180), and subunits of human integrins $\alpha 6$ and $\beta 4$. The Human Leukocyte Antigen (HLA) allele HLA-DQ $\beta 1^*0301$ has been reported to be associated with enhanced susceptibility to all of these subsets. Sera of patients with the four subsets are characterized by the presence of anti-Basement Membrane Zone (anti-BMZ) antibodies. In this manuscript, we present a model in which relevant portions of the four different antigens involved in pemphigoid have potential sites that could be presented by an antigen presenting cell (APC) in conjunction with DQ $\beta 1^*0301$ to a T cell receptor to initiate the process that results in anti-BMZ antibody production. Thus, this model provides a hypothetical computer-based mechanism to explain how a single HLA allele can be associated with the production of antibodies to four different antigens that result in four different subsets of a disease with four different clinical profiles and prognoses.

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1. Introduction

Pemphigoid (Pg) is a potentially fatal subepidermal blistering autoimmune disease. The majority of the patients are elderly [1]. Pg has

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- ★ This manuscript had not been previously presented.
- * Corresponding author. Tel.: +1 617 738 1040; fax: +1 617 754 6434. *E-mail addresses*: parecheg@med.ucm.es (P. Reche), arahmedmd@msn.com

two major phenotypes, Bullous Pemphigoid (BP) and Mucous Membrane Pemphigoid (MMP), also referred to as Cicatricial Pemphigoid (CP) [2].

BP characteristically affects elderly patients who present with large tense bullae on the entire skin and frequently the extremities [3,4]. Oral involvement is infrequently observed [3,4]. Pruritus may be significant [3]. The blisters rupture easily leaving large denuded surfaces, which can be easily infected since the blister fluid has a composition very similar to serum [5,6]. The mortality rate can vary from 19 to 30% [4]. Lesions of BP heal without scarring, but tend to leave post-inflammatory hypo- or hyper-pigmented macules [4].

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MMP affects the mucous membranes of the oral cavity, conjunctiva, nose, esophagus, pharynx, larynx, genitalia, anal canal, and the skin [3,6–11]. The lesions with MMP, upon healing, result in irreversible scarring [3,6–11]. This scarring can have catastrophic and very significant influences on the patients' quality of life. Scarring of the larynx can result in sudden asphyxiation, scarring of the esophagus requires repeated dilatation, and scarring of the anal canal, penile, and vaginal mucosa can significantly affect activities of daily living [6].

There are two subsets of MMP that deserve special mention because of the striking differences in their clinical presentation and prognosis. When MMP or CP involves predominantly or exclusively the conjunctival mucosa, it is referred to as Ocular Cicatricial Pemphigoid (OCP) [4]. The most concerning aspect of ocular involvement is that it can lead to blindness in spite of the most aggressive immunosuppressive treatment [12]. Oral Pemphigoid (OP) is that subset of MMP where the disease process is limited only to the oral cavity, and usually does not involve any other mucosa [13]. While OP is usually not fatal, eating, swallowing, and maintaining adequate nutritional levels can be both challenging and difficult [6].

The hallmark of both BP and MMP (including the subsets) is that these patients have circulating antibodies to molecules in the Basement Membrane Zone (BMZ) of the skin or the mucosal tissues [4]. These antibodies may be detected by Indirect Immunofluorescence (IIF) using a variety of substrates, the most common of which is monkey esophagus [4]. The histology of lesions from both variants of Pg shows a subepidermal vesicle with a dermal infiltrate that may be eosinophilic, neutrophilic, or mixed [2,11,14]. Direct Immunofluorescence studies of perilesional tissue in both variants demonstrate deposition of Immunoglobulin (Ig) G and/or complement along the BMZ [2,8,11].

There are two major target antigens in BP, Bullous Pemphigoid Antigen 1 (BPAG1, also known as BP230) and Bullous Pemphigoid Antigen 2 (BPAG2, also known as BP180) [2,14,15]. The major target of the autoantibody in OP is subunits of human integrin $\alpha 6$ [13]. In MMP and OCP, the target antigen is a subunit of human $\beta 4$ integrin [14,16–18]. Sera of patients with MMP may have autoantibodies that bind BPAG1 and BPAG2, but the levels and presence do not correlate with disease activity [19].

BPAG1 has a molecular weight of 230 kDa [14]. It is a desmoplakin located in the intracellular portion of the hemidesmosome complex [14]. Its gene is located on the short arm of chromosome six [14]. BPAG2 is a transmembrane hemidesmosome with a molecular weight of 180 kDa [14,20]. It has 15 domains that belong to the long carboxy-terminal that spans the lamina lucida, and a non-collagenous 16A (NC16A) domain, found adjacent to the transmembranous aspect of the ectodomain [14,20-24]. The NC16A is known to contain major BP antigenic epitopes [21,22,25–27]. Integrins are heterodimers of α and β subunits in combination, and serve an important function in cell adhesion [16]. The α 6 β 4 heterodimer is found in the hemidesmosomes of skin and mucous membranes [16]. The 120 kDa α 6 integrin has been shown to be the target antigen in OP [13]. The titers of antibodies to $\alpha 6$ subunit correlate with disease severity and activity in patients with OP [28]. The α 6 subunit contains 1073 amino acids [29]. Antibodies to the β 4 integrin subunit correlate with disease severity and activity in the sera of MMP and OCP patients [30].

Many investigators have demonstrated that in patients with BP and all the clinical variants of MMP, there is an increased susceptibility to the disease associated with the HLA- DQ β 1*0301 allele [31–34]. Moreover, reports have shown T cell and antibody binding sites in BPAG1, BPAG2, α 6, and β 4 in patients with Pg [18,20,22,26,29].

The aim of this study was to determine if there existed a possible molecular basis for a single HLA allele binding all four different antigens involved in BP and MMP and presenting them to antigenspecific T and B cells, leading to the production of four distinct antibodies to BMZ, and four distinct clinical phenotypes of pemphigoid.

2. Methods

2.1. Patients

The patients were seen at the Center for Blistering Diseases (CBD) in Boston, MA. 21 patients with BP, 100 patients with MMP and OCP, and 22 patients with OP were enrolled in this study. Some of these patients have been previously reported [31,34–36]. This study was approved by the Institutional Review Board (IRB).

2.1.1. Inclusion criteria

To be included in this study, the patients had to fulfill the following criteria:

2.1.1.1. Clinical profile.

- A. Patients with BP had large tense blisters present on the skin and no mucosal disease.
- B. Patients with Oral Pemphigoid had erosions on the gingival and other sites in the oral cavity but no disease in any other mucosal tissues or the skin on long-term follow-up (minimum three years).
- C. Patients with OCP had scarring in the conjunctiva with symblepharon and ectropion. Some had scarring of the conjunctiva and decreased visual acuity.
- D. Patients with MMP had erosive lesions in the oral cavity, pharynx, larynx, esophagus, genitalia, and anal canal. 32% of them had cutaneous involvement.

2.1.1.2. Histology. Biopsy of a fresh lesion demonstrated a subepidermal or subepithelial blister with a mixed cell infiltrate in the dermis or submucosa.

2.1.1.3. Immunopathology. Direct immunofluorescence of perilesional skin or mucosal tissue demonstrated the presence of IgG and complement along the BMZ in a homogenous linear smooth pattern.

2.1.1.4. Serological studies.

- A. In patients with BP, antibodies to BPAG1 and BPAG2 were determined by a commercially available enzyme-linked immunosorbent assay (ELISA) [37,38].
- B. In patients with OP, antibodies to α 6 integrin (105 kDa protein) determined by an immunoblot assay using bovine gingival lysate as substrate [13]. The positive control was GoH3 monoclonal antibody [13] and sera of a patient with active pemphigus vulgaris. The negative control was 25 normal human serum.
- C. In patients with OCP and MMP, antibodies to β4 integrin (205 kDa protein) were determined by an immunoblot assay using bovine gingival lysate as substrate [17,30]. The positive control was UM-SCC-20 monoclonal antibody [16] and sera of a patient with active pemphigus vulgaris. The negative control was 25 normal human serum.

2.1.1.5. MHC class II typing. High resolution HLA-MHC II typing was done by site polymerase chain reaction with sequence specific primers (PCR-SSP) [39] on DNA of each patient obtained from peripheral blood.

2.2. Determination of T cell epitopes in relevant antigens

A theoretical computer model was used to predict antigen binding sites for HLA class II in the DQ β 1*0301 allele. T cell immune responses are elicited upon the recognition of peptide-antigens bound to HLA Class II molecules. Therefore, T cell epitopes may be surmised through the prediction of antigen-HLA binding [40]. Here, we have used the RANKPEP server (http://imed.med.ucm.es/Tools/rankpep.html), to predict potential T cell epitopes within BP180, BP230, and human Integrin α 6 and β 4 [41–43] that are restricted by HLA-DQ7, the predominant HLA II molecule whose β chain is DQ β 1*0301. The

NP_000485.3 alpha 1 type XVII collagen.

Bullous pemphigoid autoantigen BP180 Epitopes predicted to bind to HLA-DO7(DOB*0301) are represented in yellow.

1 mdvtkknkrdgtevterivtetvttrltslppkggtsngyaktaslgggsrlekqslthg
61 ssgyinstgstrghastss <mark>yrrahspas</mark> tlpnspgstferkthvtrhayegsssgnsspe
121 yprkefassstrgrsqtreseirvrlqsaspstrwtelddvkrllkgsrsasvsptrnss
181 ntlpipkkgtvetkivta <mark>ssqsvsgty</mark> dat <mark>ildanlpsh</mark> vwsstlpagssmgtyhnnmtt
241 qsssllntnaysagsvfgvpnnmascsptlhpglstsssvfgmqnnlapslttlshgttt
301 tstaygvkknmpqspaavntgvstsaacttsvqsddllhkdckflilekdntpakkemel
361 limtkdsgkvftaspasiaatsfsedtlkkekqaaynadsglkaeangdlktvstkgktt
421 tadihsygssggggsgggggggggggggggggggggggggggg
481 llfglialaeevrklkarvdeler <mark>irrsilpyg</mark> dsmdriekdrlqgmapaagadldkigl
541 hsdsqeelwmfvrkklmmeqengnlrgspqpkqdmqspqpkqdrqfpqtpqipqplqhpq
601 pqgpkgqkgsvgdpgmegpmgqrgregpmgprgeagppgsgekgergaagepgphgppgv
661 pgsvgpkgssgspgpqgppgpvglqglrgevglpgvkgdkgpmgppgpkgdqgekgprgl
721 tgepgmrglpgavgepgakgamgpagpdghqgprgeqgltgmpg <mark>irgppgpsg</mark> dpgkpgl
781 tgpqgpqglpgtpgrpgikgepgapgkivtsegssmltvpgppgppgamgppgppgapgp
841 <mark>agpaglpgh</mark> qevlnlqgppgppgprgppgpsipgppgprgppgeglpgppgpgsflsns
901 etflsgppgppgppgpkgdqgppgprghqgeqglpgfstsgsssfglnlqgppgppgpg
961 pkgdkgdpgvpgalgipsgpseggssstmyvsgppgppgppgppgsisssgqeiqqyise
1021 ymqsdsirsylsgvqgppgppgppgpvttitget <mark>fdyselash</mark> vvsylrtsgygvslfss
1081 sissedilavlqrddvrqylrqylmgprgppgpgasgdgsllsldyaelssrilsymss
1141 sgisiglpgppgppglpgtsyeellsllrgsefrgivgppgppgppgipgn <mark>vwssisved</mark>
1201 lssylhtaglsfipgppgppgppgppgpzgplatyaaensdsfrselisyltspdvr
1261 sfivgppgppgppgppgdsrll <mark>stdashsrg</mark> ssssshsssvrrgssysssmstggggag <mark>s</mark>
1321 lgaggafgeaagdrgpygtdigpgggygaaaeggmyagnggllgadfagdldynelavrv
1381 sesmqrqgllqgmaytvqgppgqpgpqgppgiskvfsaysnvtadlmdffqtygaiqgpp
1441 gqkgemgtpgpkgdrgpagppghpgppgprghkgekgdkgdqvyagrrrrrsiavkp

Bullous pemphigoid autoantigen BP180 binding to HLA-DQ7(DQβ*0301)

Consensus sequence: IWHAVHAWH Optimal Score (%OPT.): 45.671 Binding Threshold: 11.70 All rows highlighted in red represent predicted binders

binders								
RANK	POS.	N	SEQUENCE	С	MW (Da)	SCORE	% OPT.	
1	505	LER	IRRSILPYG	DSM	1056.29	19.68	43.09 %	
2	1055	GET	FDYSELASH	VVS	1050.11	19.381	42.44 %	
3	1283	RLL	STDASHSRG	SSS	898.89	16.697	36.56 %	
4	211	DAT	ILDANLPSH	VWS	961.09	16.398	35.90 %	
5	841	PGP	AGPAGLPGH	QEV	757.85	16.234	35.55 %	
6	220	PSH	VWSSTLPAG	SSM	876.01	15.634	34.23 %	
7	1320	GAG	SLGAGGAFG	EAA	717.78	13.849	30.32 %	
8	765	MPG	IRGPPGPSG	DPG	818.94	13.625	29.83 %	
9	199	VTA	SSQSVSGTY	DAT	896.91	13.531	29.63 %	
10	247	SLL	NTNAYSAGS	VFG	865.85	13.391	29.32 %	
11	80	TSS	YRRAHSPAS	TLP	1026.14	12.928	28.31 %	
12	1201	VED	LSSYLHTAG	LSF	930.03	12.403	27.16 %	
13	423	TTA	DIHSYGSSG	GGG	903.91	12.396	27.14 %	
14	837	PPG	APGPAGPAG	LPG	675.75	11.938	26.14 %	

Fig. 1. Binding sites in Bullous Pemphigoid Antigen 2 (BP180) for DQ7(DQ β 1*0301). Bullous Pemphigoid Antigen 2 (BP180) peptide antigens predicted to bind to HLA-DQ7 (DQ β 1*0301) are shown in yellow. All shown peptides exceed the Binding Threshold (see Methods). POS. = Position; N = N-terminal; C = C-terminal; MW = Molecular Weight (Daltons); %OPT. = Optimal Score.

prediction of peptide-binding to HLA is carried out using Position Specific Scoring Matrices (PSSMs) derived from peptides that are known to bind to the relevant HLA molecule. In this study, if peptides received a score higher than the Binding Threshold (BT), then they were considered to bind HLA-DQ7 (DQB1*0301). Further details are described by Reche et al. [41–43].

3. Results

3.1. Human Leukocyte Antigen (HLA) Class II Gene Associations with Pg

- A. HLA Class II Genes in Patients with BP: HLA Class II gene associations were studied in 21 patients with BP. The results show a statistically significant association with DQ β 1*0301 (P<0.05) [31].
- B. HLA Class II Genes in Patients with MMP and OCP: HLA Class II gene associations were studied in 100 patients with MMP and OCP. The results show a statistically significant association with $DQ\beta 1^*0301$ (P<0.05) [34–36].
- C. HLA Class II Genes in Patients with OP: HLA Class II gene associations were studied in 22 patients with OP. The results show a statistically significant association with $DQ\beta1^*0301$ (P<0.05) [34].

3.2. Serological studies

Serological studies in patients with BP included detection of antibodies to BPAG1 and BPAG2 by an ELISA. Based on the instructions from the manufacturer, antibody levels above 9 units/mL are considered positive [37,38]. In 21 patients with BP, using the ELISA, antibodies to BPAG1 was detected in 18 patients. Antibodies to BPAG2 were detected in all 21 patients. Antibodies to both BPAG1 and BPAG2 were found in 18 patients. Using the immunoblot assay, all the patients with Oral Pemphigoid in this study had antibodies to α 6 integrin as determined by the presence of a 120 kDa band. The positive control (GoH3) monoclonocal antibody to α 6 also demonstrated an identical 120 kDa band. The sera of the pemphigus vulgaris patient bound to a 160 kDa protein (Desmoglein 3). No binding was observed by the normal human sera.

Using the immunoblot assay, all the patients with Ocular Cicatricial Pemphigoid and Mucous Membrane Pemphigoid in this study had antibodies to β 4 integrin as determined by a 205 kDa band. The positive control (UM-SCC-20) monoclonocal antibody to β 4 also demonstrated an identical 205 kDa band. The sera of the pemphigus vulgaris patient bound to a 160 kDa protein (Desmoglein 3). No binding was observed by the normal human sera.

3.3. Molecular analysis of the potential antigen binding sites on $HLA-DQ7(DQ\beta1^*0301)$

Using the computer models, we found that HLA-DQ7(DQ β 1*0301) will bind on the following peptides on BP180: amino acid 505–513, 635–643, 765–773, 841–849, 1055–1063, 1192–1210, 1283–1291, and 1320–1328 among others (Fig. 1).

Using the computer models, we found that HLA-DQ7(DQ β 1*0301) will bind on the following peptides on BP240: amino acid 209–217, 547–555, 925–933, 1295–1303, 2029–2038, and 2366–2374 among others (Fig. 2).

Using the computer model, we found that HLA-DQ7(DQ β 1*0301) will bind on the following peptides on α 6 integrin: amino acid 34–42, 341–349, 503–511, and 495–453 among others (Fig. 3).

Using the computer model, we found that HLA-DQ7(DQ β 1*0301) will bind on the following peptides on β 4 integrin: amino acid 617–625, 890–898, 1148–1156, and 1304–1312 among others (Fig. 4).

Fig. 2. Binding sites in Bullous Pemphigoid Antigen 1 (BP240) for DQ7(DQ β 1*0301). Bullous Pemphigoid Antigen 1 (BP240) peptide antigens predicted to bind to HLA-DQ7 (DQ β 1*0301) are shown in yellow. All shown peptides exceed the Binding Threshold (see Methods). POS. = Position; N = N-terminal; C = C-terminal; MW = Molecular Weight (Daltons); %OPT. = Optimal Score.

encodes the longest isoform (1), also known as dystorn-1. Epitopes predicted to bind to HLA-DQ7(DQB*0301) are represented in subw. 1 magylspasylyweegelgayedvlerykderdkvykktftkwinghlmkvrkhvndy 61 edirdyhnisilerisgetiprekpramthrighter (1) and (1) edirdyhystore 11 edgelland (1) edirdyhystore 12 edirdyhnisilerisgetiprekpramthrighter (1) edirdyhystore 12 edirdyhnisilerisgetiprekpramthrighter (1) edirdyhystore 13 edirdyhystore 14 edirdyhnisilerisgetiprekpramthrighter (1) edirdyhystore 14 edirdyhnisilerisgetiprekpramthrighter (1) edirdyhystore 14 edirdyhnisilerisgetiprekpramthrighter (1) edirdyhystore 15 edirdyhystore 15 edirdyhystore 16 edirdyhystore 16 edirdyhystore 17 edirdyhystore 18 edirdyh	<u>NP 899236.1</u> . Bullous Pemphigoid antigen 240. Dystonin isoform 1 Description : Bullous Pemphigoid antigen 240. Transcript Variant: This variant (1) represents the longest transcript and	2581 qvqillqefatrkpqyeqltaagqgilsrpgedpslrg <mark>ivkeqlaav</mark> tqkwdsltgqlsd 2641 rcdwidqaivkstqyqsllrslsdklsdldnklssslavsthpdamnqqletaqkmkqei
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2461 dteavktqveqnksfeaelkqnvnkvqelkdkltelleenpdtpeaprwkqmlteidskw		
	2461 dteavktqveqnksfeaelkqnvnkvqelkdkltelleenpdtpeaprwkqmlteidskw	3401 nrptpragsrpstakpskiptpdrkspaskidksskr

BPA240 epitopes predicted to bind HLA-DQ7(DQβ1*0301) Consensus sequence: IWHAVHAWH Optimal Score: 45.671 Binding Threshold: 11.70 All rows highlighted in red represent predicted binders. RANK POS. SEQUENCE С MW (Da) SCORE % OPT. Ν 2505 PAK AIAAVKSGG AVL 754.88 20.494 44.87 % 2 4035 KDI IDDLVKSGH KIM 965.07 19.992 43.77 % 17.945 39.29 % 3 5650 TSV **SSOAAOAAS** POV 801.82 4 3969 VEE IDAAILRSQ QFD 968.13 17.195 37.65 % 5 299 IQW IRHHVTTMS ERT 1063.23 15.434 33.79 % 3858 LARRLHSTH 1072.24 15.425 33 77 % 6 ALO EEL 7 1543 LKD AEKAGKPPF SKQ 926.09 15.061 32.98 % 8 YRDTYHPLD DWI 1363 14.818 LKY 1161.25 32.45 % 9 574 TSR LTPSVTPAY TPG 930.07 14.805 32.42 % 10 5722 RLP GYLSGKGFH SGE 947.06 14.776 32.35 % 11 1540 SKT LKDAEKAGK PPF 941.09 14.556 31.87 % 12 534 NSG FAQTLHPSL TSG 995.15 14.356 31.43 % 13 2376 LKE ISSHGLPSD 893.96 14.332 31.38 % KAL 15 4674 LDG ALRQAKGFH GEI 1009.18 13.651 29.89 % 19 2146 LLD ARGSLLPAK ND2 894.09 13.333 29.19 % 20 5765 RPG SRAGSKAGS RAS 801.86 13 219 28 94 % 21 526 ISG ITQSLNSGF AQT 948.04 12.997 28.46 % 22 23 5500 YYE FVAALHPNK DAY 978.16 12.924 28.30 % 3464 OGL 873.96 12.923 28.30 % IQSAAKSTS TQG 24 5084 RTS SVQALKRSA REL 941.1 12.679 27.76 % 25 TQK 952.16 27.05 % 2775 LRG **IVKEQLAAV** 12.355 26 218 QSN LANLEHAFY VAE 1059.2 12.316 26.97 % 27 5576 STV MVRVGGGWM ALD 951.19 12.215 26.75 % 28 970 WKV **ISPTGNEAM** VPS 901.0 12.06 26.41 % 30 4038 IDD LVKSGHKIM TAC 994.25 11.771 25.77 %

4. Discussion

In this study, we presented 21 patients with BP, 100 patients with MMP and OCP, and 22 patients with OP, all with a highly statistically significant association with the HLA-DQB1*0301 allele. Moreover, all 21 of the patients with BP, on serological studies, demonstrated binding of antibodies to BPAG2, and 18 of the patients with BP had antibodies to BPAG1. All the patients with OP demonstrated binding of BMZ antibodies to $\alpha 6$ integrin. Patients with MMP and OCP demonstrated binding of BMZ antibodies to B4 integrin. Patients with MMP and OCP were grouped together for two reasons. The first is that many of the OCP patients had extraocular disease [44]. The second reason is that both subsets are characterized by the presence of anti- β 4 integrin antibodies in their sera [19,30]. Moreover, a major aspect of this study was to examine whether epitopes that bind to T cell receptors in the various molecules in the BMZ, that are associated with the pathogenesis of pemphigoid, are similar in the computer model used in this study to those observed in in-vitro experiments. There is data in the literature to indicate that such models are effective in these predictions [45].

The role of BPAG1 and BPAG2 antigens in BP has been studied to understand the involvement of autoreactive T cells [20–22,26]. While most studies focus on the NC16A domain of the BPAG2 antigen, there are reports that implicate the domains other than NC16A in BPAG2 and others in BPAG1 that may be recognized by the autoreactive T cells [20,21]. After expressing the NC16A-mimicking residues 490– 562 of the BP180 antigen extracellular domain as fusion proteins with Glutathione S-transferase (GST), some studies showed that T cells react with the entire sequence, as well as sequences 490–534 and 507–534 [22,26]. Interestingly, the computer model in this manuscript also demonstrates that sequence 505–513 among others is a T cell epitope for BPAG2.

Employing a different experimental technique, Thoma-Uszynski et al. have identified seven epitopes in Bacculovirus generated proteins to which autoreactive T cells bind to BPAG2 [20]. These epitopes were present in the extracellular domain of BP180 and are as follows: AA residues 490-1465, AA 490-812, AA 467-567, AA 1048-1465, AA 1352–1465, and AA 809–1106 [20]. Another study also reported T cell reactivity against residues 804–1430, indicating epitopes within the extracellular domain beyond the NC16A domain [21]. The computer model reported in this study demonstrates that the potential binding sites within BPAG2 are the following sequences: 505-513, 635-643, 765-773, 841-849, 1055-1063, 1192-1210, 1283-1291, and 1320-1328 among others. It is note-worthy that these sequences detected by the computer model were all present in the peptides created in the baculovirus system by previous investigators. This observation, in some significant measure, validates the accuracy and utility value of the computer model.

The BPAG1 antigen appears also to have epitopes recognized by the T cells. However, the T cell response to the BPAG1 antigen is somewhat less reproducible than that of the BPAG2 antigen [20]. In these experiments, peptides BP230-N (residues 1–1307), BP230-C1 (residues 1881–2649), and BP230-C2 (residues 2077–2649), produced in the baculovirus system, stimulated autoreactive T cells in patients with BP [20]. The computer model reported in this study demonstrates that the potential binding sites within BPAG1 are the following sequences: 209–217, 547–555, 925–933, 1295–1303, 2029– 2038, and 2366–2374 among others. Of note is the observation that the T cell epitopes detected by earlier investigators were similar to those predicted by the computer model. As in the case of BPAG2, our observations in BPAG1 demonstrate similar T cell epitopes, validating the utility, value, and high degree of predictability of the computer model.

The computer model presented in this manuscript demonstrates that the relevant epitopes present in BPAG1 and BPAG2 can be presented to HLA-DQB1*0301 and then be presented to T cell receptors and thus produce the necessary T cell responses. The sites on human integrin β 4 subunit that bind to the T cell receptor in MMP or OCP have not been described. Similarly, the binding sites for the T cell receptor on integrin subunit α 6 in patients with OP have not been described. However, given that the computer model has predicted T cell receptor binding sites within BPAG1 and BPAG2, that were similar to those found in the literature [20], it can be anticipated that, in all likelihood, the T cell epitopes predicted by the computer in human α 6 and β 4 integrin subunits will be similar to those done in experimental studies using human T cells from patients with OP, MMP, and OCP.

In the literature, there are three studies on patients with BP in whom HLA-MHC II genes have been reported [46–48]. In two of these studies containing 97 patients, a statistically significant increased incidence of HLA-DQB1*0301 has been observed [46,48]. In a study of 25 Northern Chinese patients with BP, no statistically significant frequency of DQB1*0301 was observed [47]. This difference reflects the difference in the genetic background between Chinese and the patients in the other studies.

Four studies, which included 224 Caucasian patients with MMP reported a statistically significant incidence of HLA-DQB1*0301 [32,33,49,50]. Two studies on 20 Caucasian patients with OCP reported a statistically significant increased frequency of OCP with DQB1*0301 [33,51]. One study on 20 Caucasian patients with OP showed a statistically significant association with DQB1*0301 [49]. In another study of 11 Caucasian patients with OP, 64% carried DQB1*0301 allele [33]. However, the authors were unable to demonstrate a statistically significant observation primarily because of the use of inappropriate controls.

The cumulative literature demonstrates that the HLA Class II DQB1*0301 allele is the most frequently observed HLA Class II allele in patients with all the clinical variants or subsets of pemphigoid disease in a statistically significant correlation.

In several autoimmune diseases, studies on the immunogenetics have provided significant information in understanding their pathogenesis [52–55]. In some autoimmune diseases with multiple subtypes, a common HLA allele association has been observed. HLA-DR β 1*1501 has been shown to be associated with both benign and malignant multiple sclerosis [56]. DR β 1*0405-DQ β 1*0401/DR β 1*0802-DQ β 1*0302 genotype was shown to be associated with both acute-onset and slowly progressive type 1 diabetes, while fulminant diabetes was associated with DR β 1*0405-DQ β 1*0401/DR β 1*0405-DQ β 1*0401 genotype, as shown by a study on a Japanese population [57].

This study has provided a computer-based model, which partially explains the mechanisms by which a single HLA allele (DOB1*0301) present in all subsets of Pg patients, is capable of binding to multiple T cell epitopes within BPAG1, BPAG2, α 6 integrin, and β 4 integrin. This binding to the T cell receptor is capable of stimulating antigen specific T cells. These T cells will interact with B cells through the CD40-CD40L to produce four distinct anti-BMZ antibodies with different specificities. These four different anti-BMZ antibodies will bind to their specific target antigen and through a series of biochemical phenomena, result in the production of a subepidermal blister. In BP, such blisters will heal without scar formation. Patients with OP recover from their blistering disease without any scar formation. In contrast, patients with MMP and OCP, usually, during the healing process, develop irreversible scar formation. This scarring process can cause significant morbidity, compromise the quality of life, and in patients with ocular involvement, can result in blindness [12]. Thus, the above described process results in four distinct clinical entities, each with a different clinical outcome as demonstrated in Fig. 5. The authors recognize that while the phenotypic presentation of Pg may be influenced by different genetic factors, non-genetic factors, and soluble and insoluble mediators of immune and inflammatory processes, non-the-less HLA-DOB1*0301 may play an important role in the pathogenesis of these clinical variants of pemphigoid.

Integrin alpha chain, alpha 6 isoform a precursor [Homo sapiens].

Epitopes predicted to bind to HLA-DQ7(DQ β *0301) are represented in yellow.

MAAAGQLCLLYLSAGLLSRLGAAFNLDTREDNVIRKYGDPGSLFGFSLAMHWQLQP EDKRLLLVGAPRAEALPLQRANRTGGLYSCDITARGPCTRIEFDNDADPTSESKED OWMGVTVOSOGPGGKVVTCAHRYEKROHVNTKOESRDIFGRCYVLSONLRIEDDMD GGDWSFCDGRLRGHEKFGSCQQGVAATFTKDFHYIVFGAPGTYNWKGIVRVEQKNN TFFDMNIFEDGPYEVGGETEHDESLVPVPANSYLGFSLDSGKGIVSKDEITFVSG<mark>A</mark> PRANHSGAVVLLKRDMKSAHLLPEHIFDGEGLASSFGYDVAVVDLNKDGWQDIVIG APQYFDRDGEVGGAVYVYMNQQGRWNNVKPIRLNGTKDSMFGIAVKNIGDINQDGY PDIAVGAPYDDLGKVFIYHGSANGINTKPTQVLKGISPYFGYSIAGNMDLDRNSYP DVAVGSLSDSVTIFRSRPVINIQKTITVTPNRIDLRQKTACGAPSG<mark>ICLQVKSCFE</mark> YTANPAGYNPSISIVGTLEAEKERRKSGLSSRVQFRNQGSEPKYTQELTLKRQKQK VCMEETLWLQDNIRDKLRPIPITASVEIQEPSSRRRVNSLPEVLPILNSDEPKTAH IDVHFLKEGCGDDNVCNSNLKLEYKFCTREGNQDKFSYLPIQKGVPELVLKDQKDI ${\tt ALEITVTNSPSNPRNPTKDGDDAHEAKLIATFPDTLTYSAYRELRAFPEKQLSCVA$ NQNGSQADCELGNPFKRNSNVTFYLVLSTTEVTFDTPDLDINLKLETTSNQDNLAP ITAKAKVVIELLLSVSGVAKPSQVYFGGTVVGEQAMKSEDEVGSLIEYEFRVINLG KPLTNLGTATLNIQWPKEISNGKWLLYLVKVESKGLEKVTCEPQKEINSLNLTESH NSRKKREITEKOIDDNRKFSLFAERKYOTLNCSVNVNCVNIRCPLRGLDSKASLIL RSRLWNSTFLEEYSKLNYLDILMRAFIDVTAAAENIRLPNAGTQVRVTVFPSKTVA QYSGVPWWIILVAILAGILMLALLVFILWKCGFFKRSRYDD<mark>SVPRYHAVR</mark>IRKEER EIKDEKYIDNLEKKOWITKWNENESYS

Alpha 6 Integrin binding to DQ7(DQ β *0301)

Consensus sequence: IWHAVHAWH Optimal Score: 45.671 Binding Threshold: 11.70 All rows highlighted in red represent predicted binders

RANK	POS.	Ν	SEQUENCE	С	MW (Da)	SCORE	% OPT.
1	34	DNV	IRKYGDPGS	LFG	974.09	14.91	32.65 %
2	341	PQY	FDRDGEVGG	AVY	932.95	14.743	32.28 %
3	503	KSC	FEYTANPAG	YNP	951.01	14.403	31.54 %
4	495	PSG	ICLQVKSCF	EYT	1022.29	14.108	30.89 %
5	1050	YDD	SVPRYHAVR	IRK	1066.24	12.743	27.90 %
6	280	VSG	APRANHSGA	VVL	861.92	12.698	27.80 %
7	174	DWS	FCDGRLRGH	EKF	1042.19	12.271	26.87 %

Fig. 3. Binding sites in human integrin α 6 for DQ7(DQ β 1*0301). α 6 peptide antigens predicted to bind to HLA-DQ7(DQ β 1*0301) are shown in yellow. All shown peptides exceed the Binding Threshold (see Methods). POS. = Position; N = N-terminal; C = C-terminal; MW = Molecular Weight (Daltons); %OPT. = Optimal Score.

Epitopes predicted to bind are highlighted in the sequence as yellow.
<u>NP 000204.3</u> Integrin beta 4 isoform 1 precursor Description
Transcript Variant: This variant (1) encodes the longest isoform (1).
1 magprpspwarllaalisvslsgtlanrckkapvksctecvrvdkdcayctdemfrdrr 61 cntqaellaagcqresivvmessfqiteetqidttlrsqmspqglrvrlpgeerhfel 121 evfeplespvdlyilmdfsnsmsddldnlkkmgqnlarvlsqttsdytigfgkfvdkvsv 181 pqtdmrpeklkepwpnsdppfsfknvisltedvdefmklqgerisgnldapeggfdail 241 qtavctrdigwrpdsthllvfstesafhyeadganvlagimsrnderchldttgtytqyr 301 tqdypsyptlvrllakhniipifavtnysysyyeklht/fpvsslgvlqedssnivelle 361 eafnrirsnldiraldsprglrtevtskmfqktrtgsfhirrgevgjvqvqlralehvdg 421 thvcqlpedqkgnihlkpsfsdglkmdagiicdvctcelqkevrsarcsfngdfvcgqcv 481 csegwsgqtcncstgslsdiqpclregedkpcsgrgecqcghcvcygegryegqfceydn 541 fqcprtsgflcndrgrcsmgqcvcepgwtgpscdcplsnatcidsnggicngrghcecgr 601 chchqslytdticeinysaihpglcedlrscvqcawgtgekkgrtceenfkvkmvde 661 lkraeevvrcsfrdeddctysytmegdgapgnstvlvhkkkdcppgsfwwlipIlll 721 llpllalllllcvkycacckaclallpccnrghmvgfkedhymlrenlmasdhldtpmlr 781 sgnlkgrdvrvkkvtnmgrpgfatnaasinptelvpyglslrlarlctenllkpdtrec 841 aqlrqeveenlnevyrqisgvklqqtkfrqqpnagkkqdhtivdtvlmaprsakpallk 901 tlekqveqrafhdlkvapgyytltadqdargmvefqegvelvdvrvplfirpedddekql 961 lveaidvpagtatlgrrlvnitiikeqardvvsfeqpefsvsrgdqvaripvirrvldgg 1021 kqusyrtqdgtaqgnrdyipvegellfqpgeawkelqvkllelqevdsllrgrqvrfh 1081 vqlsnpkfgahlgqphsttiirdpdeldrsftsqmlssqpppdglgapqnpnakaags 1141 rkihfmvlppsgkpmgyrvkywiggdseseahlldskvpsvetlnlypycdyemkvcayg 1201 aqgegpysslvscrthqevpsepgrlafnvvsstvtqlswaepaetngeitavevcyglv 1261 nddnrpigpmkkvlvdnpknrmllienlresqpyrytvkamgagggprgsatpgppge 1441 hlvngrmdfafpgstnslnrmttsaaaygthlsphvphrvlststtlrdynsltrseh 1501 shsttlprdystitsvshdsrtagvpdtptrvfsalgptslrswgqprcerplqgy 1561 sveyqllnggelnflitprsapsyrtvtfalspagegrgregvites 1621 qvhqsplcplpgsaftlypsapglvtptrvfsalgptslrswgqrcergvites 1621 qvhqsplcplpgsaftlypsaystruttsatepflvdgltlgaqhleaggsttrhvtqef 1801 vsrtitsgitstmdqqffrt

Beta 4 Integrin binding to DQ7(DQβ*0301) Consensus sequence: IWHAVHAWH Optimal Score: 45.671 Binding Threshold: 11.70 All rows highlighted in red represent predicted binders.

RANK	POS.	N	SEQUENCE	С	MW (Da)	SCORE	% OPT.
1	1208	FNW	LPPSGKPMG	YRV	865.06	20.182	44.19 %
2	1362	SQP	YRYTVKARN	GAG	1152.32	19.166	41.97 %
3	647	CEI	NYSAIHPGL	CED	953.07	19.047	41.70 %
4	932	VLM	APRSAKPAL	LK9	892.08	18.434	40.36 %
5	771	CWK	YCACCKACL	ALL	959.23	16.829	36.85 %
6	1014	EAI	DVPAGTATL	GRR	825.91	15.922	34.86 %
7	911	QTK	FRQQPNAGK	KQD	1027.15	14.639	32.05 %
9	425	EVG	IYQVQLRAL	EHV	1085.32	14.228	31.15 %
10	1421	FLM	YSDDVLRSP	SGS	1033.12	13.589	29.75 %
11	1689	FRV	RAQSQEGWG	RER	977.04	13.035	28.54 %
12	661	DLR	SCVQCQAWG	TGE	940.09	12.643	27.68 %
13	970	TLT	ADQDARGMV	EFQ	944.03	12.108	26.51 %
14	1060	RIP	VIRRVLDGG	102	966.15	11.829	25.90 %

Fig. 4. Binding sites in human integrin $\beta4$ for DQ7(DQ $\beta1^*0301$). $\beta4$ peptide antigens predicted to bind to HLA-DQ7(DQ $\beta1^*0301$) are shown in yellow. All shown peptides exceed the Binding Threshold (see Methods). POS. = Position; N = N-terminal; C = C-terminal; MW= Molecular Weight (Daltons); %OPT. = Optimal Score.

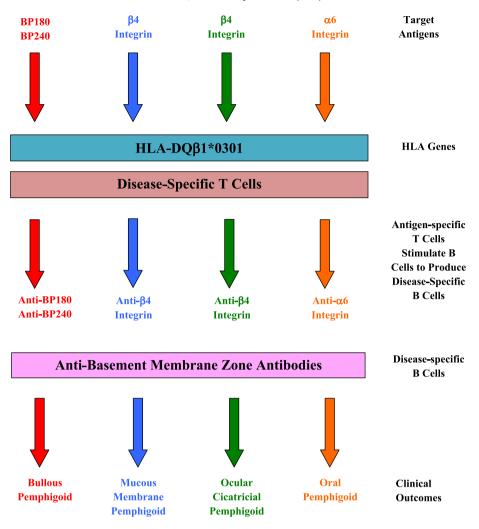


Fig. 5. Summary of proposed mechanism by which a single HLA allele (DQ₃1*0301) binds all four different pemphigoid antigens and produces four distinct phenotypes of pemphigoid disease. Variants of pemphigoid disease - bullous pemphigoid, mucous membrane pemphigoid, ocular cicatricial pemphigoid, and oral pemphigoid, all are characterized by anti-BMZ antibodies. Each of these has different antigenic targets. They all have a common HLA MHC Class II allele in high frequency associated with them, and it recognizes epitopes within each of these antigens. Resultantly, a different antibody is produced in each variant producing a different phenotype. (BP240=Bullous Pemphigoid Antigen 1; BP180=Bullous Pemphigoid Antigen 2; HLA=Human Leukocyte Antigen).

Take-home messages

- Pemphigoid has four clinically dissimilar variants. These are BP, MMP, OCP, and OP.
- All variants produce anti-BMZ antibodies, each targeting different antigens.
- The antigens: BPAG1 and BPAG2 in BP; β4 integrin in MMP and OCP; α6 integrin in OP.
- All variants have significant enhanced susceptibility associated with HLA-DQ\(\beta\)1*0301.
- T cell epitopes in the antigens bound to DQβ1*0301 by computer model.

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Boundary stones in Systemic Lupus Erythematosus: Free Light Chains

The assessment of disease activity in systemic lupus erythematosus (SLE) is a challenge of immunologists and rheumatologists. Serological parameters, such as complement C3 and C4 and anti-dsDNA antibodies, do not always correlate with clinical activity of the disease, and the search for new "biomarkers" could provide a window of opportunity for therapeutic intervention to prevent onset of irreversible damage. Recently, Aggarwal et al. have evaluated the role of free light chains (FLC) in the assessment of disease activity. These are produced in excess of heavy chains These excess FLC are released into the serum, from which they are rapidly removed by the kidneys.

The authors found that FLC was higher in SLE than in RA patients, and both were higher than healthy controls. Especially, SLE patients with high disease activity were those who showed the highest total FLC was significantly higher. Interestingly, these patients were not differentiated by IgG, C4, or dsDNA antibody levels. Nonetheless, total FLC and C3 showed moderate to strong correlation with the SLEDAI. There are several mechanisms that could explain such increase in FLC in SLE patients, such as a dysregulated or inefficient process of receptor. FLC seem to be more sensitive than IgG, C4, or dsDNA antibody as they would be unaffected by the constituent immune reactions and acute inflammatory processes.

This must be confirmed in larger case series, however, these preliminary results seem to previse a future for the usage of FLC as biomarkers of SLE activity.